

GC-MS ANALYSIS OF THE VOLATILE CONSTITUENTS FROM THE AIR-DRIED LEAVES OF *Terminalia catappa* (LINNAEUS)



Abdulrazaq O. Ogunmoye*, Oseyemi O. Olubomehin, C. O. Atewolara-Odule, Segun A. Ogundare and Sodiq T. Yussuf

Department of Chemical Sciences, Olabisi Onabanjo University, PMB 2002, Ago-Iwoye, Ogun State, Nigeria *Correspondent author: <u>omotunde.ogunmoye@oouagoiwoye.edu.ng</u>

Received:August 11, 2020Accepted:October 02, 2020Abstract:The volatile constituents of air-dried leaves (200 g) of *Terminalia catappa* growing in Nigeria was obtained by
hydrodistillation using an all glass Clevenger-type glass apparatus. The extracted oil was analyzed using a gas
chromatography coupled with mass spectrometry (GC-MS). A total of fourteen constituents' representing 74.32%
of *T. catappa* oil with a yield of 0.25% (v/w) was obtained. Hexahydrofarnesyl acetone (12.34%) was the main
constituent of the oil followed by 1,3,8-p-Menthatriene (9.38%), 1,2-dimethyl-Cyclooctene (7.30%) Undecane
(6.73%), Trans-geranylacetone (6.02%) and 4-ethyl-m-xylene (5.83%). The other constituents' were; Cis-13-
Octadecenal (4.83%), Trans-2-Decenol (4.28%), 3-Eicosene, (E)- (4.17%), 1,7-Hexadecadiene (3.47%), m-Propyl-
toluene (3.01%), Neophytadiene (2.56%) trans-β-Ionone (2.53%) and 2-ethyl-p-xylene (1.87%). The common
classes of compounds in the leave oil are; Hydrocarbons (21.67%), Oxygenated Sesquiterpenes (14.87%), Benzene
derivatives (10.71%), Monoterpenes (9.38%), Oxygenated hydrocarbons (9.11%), Oxygenated monoterpene
(6.02%) and Sesquiterpenes (2.56%).

Keywords: Combretaceae, essential oil, hexafarnesyl acetone and *Terminalia catappa*

Introduction

The genus Terminalia includes 200 species, some of them with economic importance as ornamentals and timber yielding plants (Mabberley, 2008). Terminalia catappa L., a member of the Combretaceae family is a perennial tree species found in almost all the regions of the Country as it thrives well in the tropics. It is commonly called tropical almond, wild almond, Indian almond, sea almond, beach almond and Malabar almond (Untwal and Kondawar, 2006; Orwa et al., 2009). It is mainly found in the southern part of Nigeria, especially in the south-east where they are usually planted for provision of shade and ornamental purposes (Ezeokonkwo and Dodson, 2004; Agu and Menkiti, 2017). It is widely planted throughout the tropics, especially along sandy seashores, for shade, ornamental purposes, sand-dune stabilizer and edible nuts (Brown and Cooprider, 2013). Furnitures and interior building materials are derived from the hardword of the Plant (Lex and Barry, 2006).

Tropical almond has a characteristic 'pagoda' shape because it sends out a single stem from the top centre (Chen *et al.*, 2000). The fruit has an endocarp, which contains an edible oily seed that tastes like almond. The dried raw seeds of tropical almond are highly relished by children in India, Malaysia and Nigeria (Sosulski *et al.*, 1998; Henn *et al.*, 2014). Fruits are produced from about 3 years of age, and the nutritious, tasty seed kernels may be eaten immediately after extraction (Mohale *et al.*, 2009).

The leaves of *Terminalia catappa* are used as a maturant and emollient; the juice is used in the preparation of ointment for scabies, leprosy and other cutaneous diseases (Nair and Chanda, 2008). It has also been investigated for its medicinal activities. These include the *in vitro* and *in vivo* antimetastic effects (Chen *et al.*, 2007), antidiabetic activities (Rao and Nammi, 2006), actinociceptive activity, antiparasitic, antibacterial, antifungal, antimicrobial (Elizabeth, 2005; Nair and Chanda, 2008; Rajarajan *et al.*, 2010), antioxidant (Ko *et al.*, 2002; Mety and Mathad, 2011), anticancer properties (Chu *et al.*, 2007), hepatitis and liver-related diseases in Taiwan (Lin and Kan, 1990).

In addition, some of the phytochemicals identified from *T. catappa* leaves extracts includes; triterpenic acids responsible for the antiinflammatory activity (Fan *et al.*, 2004), hydrolysable tannins, such as punicalagin, punicalin, terflavins A and B, tergallagin, tercatain, chebulagic acid, geraniin, granatin B, and corilagin, but no caffeine (Tanaka *et*

al., 1986). Also identified are Six phenolic compounds; phydroxybenzoic acid, 4-hydroxyphenylpropionic acid, mcoumaric acid, 3,4-dihydroxybenzoic acid, p-coumaric acid and gallic acid (Chyau et al., 2006). Furthermore, apigenin 6-C-(2-O-galloyl)-â-Dglucopyranoside, apigenin 8-C-(2-Ogalloyl)- â -D-glucopyranoside, glycosides, sovitexin, vitexin, isoorientin, and rutin, were isolated from the dried fallen leaves of Terminalia catappa (Yun-Lian et al., 2000). The Leaf also contains 1-degalloyl-eugeniin, 2,3-(4,4',5,5',6,6'hexahydroxy-diphenoyl)-glucose, chebulagic-acid, corilagin, gentisic-acid, geraniin, granatin-b, kaempferol, punicalagin, punicalin, quercetin, tercatain, terflavin-a, terflavin-b. tergallagin (Duke, 2008).

The chemical compositions of some essential oils from some other Terminalia species have been reported. For example the water-distilled essential oil of *Terminalia bentzoë* from the Island of Rodrigues analyzed by GC and GC/MS gave Twenty-eight constituents which was dominated by citronellyl acetate (64.87%) (Gurib-Fakim and Demarne, 1994).

Furthermore, the composition of essential oil from the fruit oil of *T. chebula* obtained from India was found to consist mainly of palmitic acid (35.7%), furfural (26.8%) and phenylacetaldehyde (13.1%) (Naik *et al.*, 2010). In addition, the main compounds of the oil from *Terminalia ivorensis* flower from Northern Nigeria were δ -3-carene (29.4%) and α -pinene (20.9%) (Ogunwande *et al.*, 2019).

Although, the essential oil of the different parts (fruits, leaves, nuts) of *T. catappa* has been analyzed with some of the following major phytochemicals identified; α -farnesene (21.3%), octadedecane (11.7%), hexadecanoic acid (9.5%), dibutyl phthalate (9.1%), 1,2,3-trimethoxy-5-(2-propenyl)-benzene (6.6%), neoisothujol (5.8%), 1,2,4-trimethoxy-5-(1-propenyl)-benzene (4.5%) 6,10,14-trimethyl-2-pentadecanoic, 1-(2,3,6-trimethyl phenyl)-(E)-3-buten-2-one, geranyl acetone, hexadecanoic acid (21.0%) and 2-ethyl-3,6-dimtheylpyrazine (19.2%), (Z)-phytol (41.2%), fatty acid palmitic acid (11.0%), and the (E)-nerolidol (4.7%) (Moronkola and Ekundayo, 2000; Wang *et al.*, 2000; Lasekan *et al.*, 2012; Owolabi *et al.*, 2013).

In continuation of our research into the volatile oil components of some poorly studied plants grown in Nigeria, we report in this paper the essential oil constituents of the airdried leaves of *T. catappa* as our contribution towards increasing the available literature on the plant due to limited work on the essential oil of the leave.

Materials and Methods

Sample collection and identification

The leaves of *Terminalia catappa* were collected from a tree sample in Olabisi Onabanjo University (permanent site), Ago-Iwoye, Ogun State, Nigeria. Identification and authentication was done at the Herbarium of Forestry Research Institute of Nigeria (FRIN), Ibadan, Nigeria where a voucher specimen with herbarium no. FHI 110462 was deposited.

Extraction of essential oil

The pulverized air-dried leaves of *T. catappa* (200 g) were subjected to hydrodistillation in an all glass Clevenger-type all glass apparatus for 3 h in accordance with established procedure (British Pharmacopoeia, 1980). The oil collected was preserved in a sample tube and stored in a

Refrigerator until analysis.

Gas chromatography-mass spectrometry (GC/MS) analysis of the leaf oil

GC-MS analysis of the oil was done on an Agilent model 7890A gas chromatograph equipped with a FID and fitted with a fused silica capillary HP-5 MS column (30 m x 0.32 mm id, film thickness 0.25 μ m). The oven temperature was programmed from 80 – 240°C at the rate of 8°C/min. The ion source was set at 240 °C and electron ionization at 70 eV. Helium was used as the carrier gas at a flow rate of 2 mL/min. Scanning range was 35-425 amu. Diluted oil in n-hexane (1.0 μ L) was injected into the GC/MS spectrometer.

Identification of the constituents of the leaf oil

The individual components of the oil were identified on the basis of their retention indices (RI) determined by co-injection with reference to a homologous series of n-alkanes, under identical experimental conditions. Further identification was performed by comparison of their mass spectra with those from National Institute of Standards and Technology NIST (Data base 69) and the home-made MS library built up from pure substances and components of known essential oils, as well as by comparison of their retention indices with literature values (Adams, 2007).

Results and Discussion

The essential oil (0.25% v/w), obtained was cloudy light yellow with a strong odour. The result of the chromatographic analysis done on the essential oil is shown in Fig. 1 while the identities of the constituents as well as their percentage composition are shown in Table 1. Fourteen constituents were identified representing 74.32% of the oil from T. catappa with the following components; Hexahydrofarnesyl acetone (12.34%) as the major constituent, followed by 1,3,8-p-Menthatriene (9.38%), 1,2-dimethyl-Cyclooctene (7.30%), Undecane (6.73%) Trans-geranylacetone (6.02%) and 4-ethylm-xylene (5.83%). The other constituents of the oil were Cis-13-Octadecenal (4.83%), Trans-2-Decenol (4.28%), 3-Eicosene, (E)- (4.17%), 1,7-Hexadecadiene (3.47%), m-Propyl-toluene (3.01%), Neophytadiene (2.56%) trans-β-Ionone (2.53%) and 2-ethyl-p-xylene (1.87%). Hydrocarbons (21.61%) were the most abundant class of compounds in the oil composition while the others are Oxygenated Sesquiterpenes (14.87%), Benzene derivatives (10.71%), Monoterpenes (9.38%), Oxygenated hydrocarbons (9.11%), Oxygenated monoterpene (6.02%) and Sesquiterpenes (2.56%).

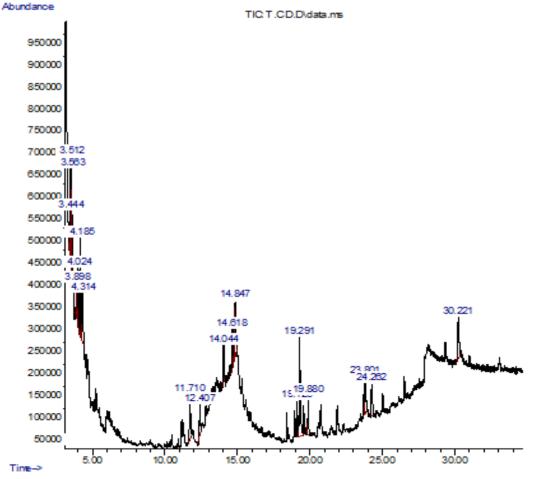


Fig. 1: Chromatogram of the air-dried leaves of Terminalia catappa essential oil components

Essential Oil Constituents	of Terminalia catappa Air-dried Leav	es
----------------------------	--------------------------------------	----

Table 1: Chemical composition of essential oil from	n the air-dried leaves of <i>Terminalia catappa</i>
---	---

S/N	Name	KOVAT index	Retention time	% Composition	Molecular formula
1	m-Propyl-toluene	981	3.442	3.01	C10H14
2	1,2-dimethyl- Cyclooctene	1000	3.510	7.30	$C_{10}H_{18}$
3	1,3,8-p-Menthatriene	1000	3.562	9.38	$C_{10}H_{14}$
4	2-ethyl-p-xylene	1000	3.899	1.87	$C_{10}H_{14}$
5	4-ethyl-m-xylene	1044	4.025	5.83	$C_{10}H_{14}$
6	Undecane	1000	4.185	6.73	$C_{11}H_{24}$
7	Trans-2-Decenol	1103	4.311	4.28	$C_{10}H_{20}O$
8	Trans-geranylacetone	1274	11.710	6.02	C13H22O
9	Trans-β-Ionone	1366	12.408	2.53	$C_{13}H_{20}O$
10	Neophytadiene	1788	19.125	2.56	$C_{20}H_{38}$
11	Hexahydrofarnesyl acetone	1912	19.291	12.34	C18H36O
12	1,7-Hexadecadiene	1800	19.881	3.47	C16H30
13	Cis-13-Octadecenal	1958	23.800	4.83	C18H34O
14	3-Eicosene, (E)-	1835	24.264	4.17	$C_{20}H_{40}$
		Toluene/Benzene derivatives		10.71	
			Hydrocarbons		
		Oxygenate	Oxygenated hydrocarbons		
		Monoterpene		9.38	
		Oxygenated monoterpene		6.02	
		Sesquiterpene		2.56	
		Oxygenated sesquiterpenes		14.87	

Hexahydrofarnesyl acetone, which was the major constituent present in the oil extracted in this study belong to the sesquiterpenoids class of compound and its quantitative amount of (12.34%) in the oil is noteworthy since it has not been previously reported to be a major compound of *T. catappa* (Owolabi *et al.*, 2013).

A comparison of the present result with previously analysed samples of *T. catappa* revealed some qualitative and quantitative variations. For example, Sixty-six constituents accounting for 100% of the composition included (*Z*)-phytol (41.2%) an acyclic diterpenoid, palmitic acid (11.0%) a fatty acid being the dominant compounds and lesser quantities of the sesquiterpenoid- (*E*)-nerolidol (4.7%), alkane hydrocarbons; heptadecane (3.0%), hexadecane (2.3%), pristane (2.2%), and phytane (2.0%) (Owolabi *et al.*, 2013).

Similarly, (*Z*)-phytol (41.2%), the main compound of *T. catappa* (Owolabi *et al.*, 2013), was not identified in this sample while the content of trans β -ionone (0.8%) was also insignificant compared to the 2.53% of this present work. It was also observed that common constituents like linalool, camphor, menthol, α - Humulene and (E)-Caryophyllene present in the leaf oil of previous study from Nigeria, were not detected in the current sample.

However, hexahydrofarnesyl acetone (12.34%) which was the major constituent, followed by 1,3,8-p-Menthatriene (9.38%), 1,2-dimethyl-Cyclooctene (7.30%) Undecane (6.73%) and 4-ethyl-m-xylene (5.83%) were not identified in previous studies (Owolabi *et al.*, 2013). Also, Hexahydrofarnesyl acetone was previously reported to exhibit a potent antimicrobial activity against gram-positive and gram-negative bacteria (Filipowicz *et al.*, 2003), antibacterial, antifungal, (Radulovic *et al.*, 2006), antimicrobial (Radulovic*et al.*, 2011), had allopathic (Razavi and Nejad-Ebrahimi, 2010) and pest control potential (Mohamed *et al.*, 1992).

Nevertheless, the presence of (5E,9E)-fernesyl acetone, geranylacetone, trans β -ionone and other saturated isomeric compounds in previously reported data are in accordance with the current study though non-quantitatively (Mau *et al.*, 2003; Owolabi *et al.*, 2013). The chemical compositions of some essential oils of the different parts of *T. catappa* from other parts of the world have been reported. The oil of *T. catappa* leaves from Taiwan was characterised by the abundance of 6,10,14-trimethyl-2-pentadecanoic, 1-(2,3,6-trimethyl phenyl)-(E)-3-buten-2-one and geranyl acetone (Wang *et al.*,

2000). The composition of the roasted *T. catappa* nut from Malaysia was dominated by Hexadecanoic acid (21.0%) and 2-ethyl-3,6-dimtheylpyrazine (19.2%) (Lasekan *et al.*, 2012). From the fore-going, there are obvious qualitative and/or quantitative compositional variations in the volatile compounds present in the different parts of *T. catappa* as well as other species from the same genus and this may be due to ecological factors, age of the plant, period of collection, handling procedure and climatic condition (Inikpi *et al.*, 2014; Rehman *et al.*, 2016; Ibanez and Blazquez, 2019).

Conclusion

The essential oil of the leaves of *T.catappa* investigated revealed the presence of fourteen constituents as determined by GC-MS analysis constituting 74.32% of the total oil composition. The presence of hexahydrofarnesyl acetone as the major constituent in the oils justifies the traditional use of the plant in treating pains, headaches and as an antimicrobial agent. Also, some of the identified chemical constituents from the plant are new additions to the chemical data base for the plant.

Acknowledgment

Authors are grateful to Miss Oyebade, Doyinsola Christianah for the assistance in the extraction of the oil sample.

Conflict of Interest

Authors declare that there is no conflict of interest related to this study.

References

- Adams RP 2007. Identification of Essential Oil Components by Gas Chromatography/Mass Spectrometry, 4th Ed. Allured Publishing Corp, Carol Stream, Illinois.
- Agu CM & Menkiti MC 2017. Effects of natural antioxidants on the essential properties of modified *Terminalia catappa* L. kernel oil: A possible substitute for mineral oil transformer fluid. *Biofuels*.
- British Pharmacopoeia 1980. Her Majesty;s Stationary Office, Atlantic House, Holborn Viaduct, London, England Vol. II, pp. 109.
- Brown SH & Cooprider K 2013. *Terminalia catappa*. U.S. department of Agriculture, Cooperative Extension Service, University of Florida, IFAS, Lee County Extension, Florida. <u>http://lee.ifsa.ufl.edu/Hort/GardenPubsAZ/tropical_almond.pdf</u>

- Chen CS, Fa YS, Jung LS, Hsien KW, Zin CY & Shou HY 2007. In vitro and in vivo antimetastatic effects of *Terminalia catappa* L. *Food and Chemical Toxicology*, 45(7): 1194-1201.
- Chen PS, Li JH, Liu TY & Lin TC 2000. Folk medicine *Terminalia catappa* and its major tannin component, punicalagin are effective against bleomycin-induced genotoxicity in Chinese hamster ovary cells. *Cancer Letters*, 152: 115–122.
- Chu SC, Yang SE, Liu SJ, Kuo WH & Chang YZ 2007. *Terminalia catappa* L. leaves on lung cancer Cells. *Food and Chemical Toxicology*, 45(7): 1194-1201.
- Chyau CC, Ko PT & Mau JL 2006. Antioxidant properties of aqueous extract from *Terminalia catappa* leaves. *Food Sci. Technol.*, (LWT) 39: 1099-1108.
- Duke JA 2008. Phytochemical and Ethnobotanical Databases (online database).
- Elizabeth KM 2005. Antimicrobial activity of *Terminalia* bellerica. Indian J. Clin. Biochem., 20(2): 150-153.
- Ezeokonkwo CA & Dodson WL 2004. The potential of *Terminalia catappa* (tropical almond) seed as a source of dietary protein. *J. Food Qual.*, 27(3): 207–219.
- Fan YM, Xu LZ, Gao J, Wang Y, Tang XH, Zhao XN & Zhang ZX 2004. Phytochemical and antiinflammatory studies on *Terminalia catappa*. *Fitoterapia*, 75: 253–260.
- Filipowicz N, Kaminski M & Kurlenda J 2003. Antibacterial and antifungal activity of juniper berry oil and its selected components. *Phytother. Res.*, 17: 227–231.
- Gurib-Fakim A & Demarne F 1994. Essential oil of *Terminalia* bentzoë (L.) L. f. subsp. Rodriguesensis Wickens., Journal of Essential Oil Research, 6(5): 533-534.
- Henn JJ, McCoy MB & Vaughan CS 2014. Beach almond (*Terminaliacatappa*, Combretaceae) seed production and predation by scarlet macaws (Aramacao) and variegated squirrels (*Sciurusvariegatoides*). *Revista De Biologia Tropical*, 62(3): 929–38.
- Ibanez MD & Blazquez MA 2019. Essential oils: Quality indicators of spices in supermarkets, Nereis. Interdisc. Ibero-Ame. J. Methods, Modelling and Simulation Marzo, 11: 39-50.
- Inikpi E, Lawal OA, Ogunmoye AO and Ogunwande IA 2014. Volatile composition of the floral essential oil of *Hibiscus sabdariffa* L. from Nigeria. Am. J. Essential Oils and Nat. Prod., 2(2): 4-7.
- Ko TF, Weng YM & Chiou RYY 2002. Squalene content and antioxidant activity of *Terminalia catappa* leaves and seeds. J. Agric. Food Chem., 50: 5343-5348.
- Lasekan O, Alfi K & Abbas KA 2012. Volatile compounds of roasted and steamed Malaysian tropical almond nut (*Terminalia catappa* L.). Int. J. Food Prop., 15(5): 1120-1132.
- Lex AJT & Barry E 2006 Terminalia catappa (tropical almond) Combretacea (combretum family) *Species Profiles for Pacific Island Agroforestry* Ver., 2(2): 2.
- Lin CC & Kan WS 1990. Medicinal plants used for the treatment of hepatitis in Taiwan. Am. J. Chinese Med., 18: 35–43.
- Mabberley DJ 2008. Mabberly's Plant-Book, 3rd Ed. Cambridge Univ. Press, Cambridge, UK, pp. 846-847.
- Mau JL, Ko PT & Chyau CC 2003. Aroma characterization and antioxidant activity of supercritical carbon dioxide extracts from *Terminalia catappa* leaves. *Food Res. Int.*, 36: 97-104.
- Mety SS & Mathad P 2011. Antioxidative and free radical scavenging activities of *Terminalia* species. *Int. Res. J. Biotech.*, 2(5): 119-127.
- Mohale DS, Dewani AP, Chandewar AV, Khadse CD, Tripathi AS & Agrawal SS 2009. Brief review on

medicinal potential of *Terminalia catappa*. *J.of Herbal Med. and Toxicol.*, 3(1): 7-11.

- Mohamed MA, Quisenberry SS & Moellenbeck DJ 1992. 6,10,14- Trimethylpentadecan-2-one: A Bermuda grass phagostimulant to fall armyworm (Lepidoptera: Noctuidae). J. Chem. Ecol., 18(4): 673–682.
- Moronkola DO & Ekundayo O 2000. Chemical constituents in the fruit essential oil of *Terminalia catappa* Linn (almond fruits). J. Trop. For. Res., 16(1): 72-79.
- Naik DG, Puntambekar H & Anantpure P 2010. Essential oil of *Terminalia chebula* fruits as a repellent for the Indian honeybee *Apisflorea*. *Chem. Biodiver.*,7(5): 1303-1310.
- Nair R & Chanda S 2008. Antimicrobial activity of *T erminalia catappa*, manilkarazapota and piper betel leaf extract. *India J. Pharm. Sci.*, 70(30): 390-393
- Ogunwande IA, Ascrizzi R & Guido F 2019. Essential oil composition of *Terminalia ivorensis* A. Chev. flowers from northern Nigeria. *Trends in Phytochem. Res.*, 3(1): 77-82
- Orwa C, Mutua A, Kindt R, Jamnadass R & Anthony S 2009. Agroforestree Database: A Tree Reference and Selection Guide Version 4.0.
- Owolabi MS, Lawal OA, Ogunwande IA, Hauser RM, William N & Setzer WN 2013. Chemical composition of the leaf essential oil of *Terminalia catappa* L. growing in southwestern Nigeria. Am. J. Essential Oils and Nat. Prod., 1(1): 51-54.
- Radulovic N, Stojanovic G & Palic R 2006. Composition and antimicrobial activity of *Equisetum arvense* L. essential oil. *Phytother. Res.*, 20: 85–88.
- Radulovic N, Dekic M, Stojanovic-Radic ZO & Palic R 2011. Chemical composition and antimicrobial activity of the essential oils of *Geranium columbinum* L. and *G. lucidum* L. (Geraniaceae). *Turk. J. Chem.*, 35(3): 499–512.
- Rajarajan S, Asthana M & Shanthi, G 2010. The in vitro bactericidal activity of lyophilized ethanolic extract of Indian almond (*Terminalia catappa* Linn) fruit pulp on two pathogenic bacteria from subgingival plaques. *Indian J. Nat. Prod. and Resou.*, 1(4): 466-469.
- Rao NK & Nammi S 2006. Antidiabetic and renoprotective effects of the chloroform extract of *Terminalia chebula*retz, Seeds in streptozotocinduced diabetic rats. *BMC: CAM*, 6: 17-17.
- Razavi SM & Nejad-Ebrahimi S 2010. Phytochemical analysis and allelopathic activity of essential oils of *Ecballium elaterium* A. Richard growing in Iran. *Nat. Prod. Res.*, 24(18): 1704–1709.
- Rehman R, Hanif MA, Mushtaq Z and Al-Sadi AM 2016. Biosynthesis of essential oils in aromatic plants: a review. *Food Reviews International*, 32(2): 117–160.
- Sosulski FW, Abdullahi AH, Sosulski K 1988. Potential of tropical almond (*Terminaliacatappa*) fruits as a source of edible oil. La Rivista Delle Sostanze Grasse, 65: 21–23.
- Tanaka T, Nonaka GI & Nishioka I 1986. Tannins and related compounds. XLII. Isolation and characterization of four new hydrolysable tannins, terflavins A and B, ergallagin and tercatain from leaves of *Terminalia catappa L*. *Chem. and Pharmac. Bull.*, 34: 1039–1049.
- Untwal LS & Kondawar MS 2006. Use of *Terminalia catappa* fruit extract as an indicator in acid-base titration. *Indian J. Pharmac. Sci.*, 68(3): 399-401.
- Yun-Lian L, Yueh-Hsiung K, Ming-Shi S, Chien-Chih C & Jun-Chih O 2000. Flavonoid glycosides from *Terminalia* catappa L. J. Chin. Chem. Soc., 47: 253-256.
- Wang HF, Ko P, Chyau CC, Mau JL & Kao MD 2000. Composition and antioxidative activity of essential oils from *Terminalia catappa* L. leaves. *Taiwanes eJournal* of Agric. Chem. and Food Sci., 38(1): 27-35.